

## Phenol/chloroform extraction

1. Add an equal volume of phenol/chloroform to the DNA-containing solution.
2. Vortex for 15 seconds.
3. Centrifuge for 3 minutes.
4. Remove upper aqueous layer and place in a fresh eppendorf.
5. If aqueous layer is 200 $\mu$ l or less, add an equal volume of TE<sub>1</sub><sup>or H<sub>2</sub>O</sup> to the remaining phenol solution and vortex.
6. Centrifuge for 3 minutes and combine aqueous layers.

## Ethanol precipitation

1. Add one tenth volume of 3M NaAc, pH 5.2 (and 1 $\mu$ l glycogen if DNA is less than 500bp) to aqueous solution.
2. Add 2.5-3 volumes of cold 100% EtOH.
3. Place at -20°C overnight or at -70°C for 30 mins. [20 mins will be OK]
4. Centrifuge for 15 mins at 4°C then discard s/n.
5. Wash pellet with 1ml cold 70% EtOH and invert to mix.
6. Recentrifuge for 10 mins at 4°C and again discard s/n.
7. Dry pellet under vacuum then resuspend in 50 $\mu$ l/10 $\mu$ l TE. Vortex briefly and spin to bring down suspension. Store at -20°C.

Resuspend fragments in H<sub>2</sub>O